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# **MicroRNA-21 as a Potential Biomarker for Colorectal Cancer Diagnosis: A Meta-analysis**

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# *Authors' contributions*

*This work was carried out in collaboration between all authors. Author CHZ wrote the protocol and the first draft of the manuscript. Author GJY collected, processed and analyzed data. Author YT managed the literature searches. Author LY designed research and contributed to interpretation of data. Authors XFY and FRL contributed to research design, revising the paper critically and approval of the submitted and final versions. All authors read and approved the final manuscript.*

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# **ABSTRACT**

**Aims:** Colorectal cancer (CRC) occupies an important position in the morbidity and mortality constitution of malignancies. In recent years, mounting literature has reported about the upregulating expression of microRNA-21 in blood and stool of CRC patients, which suggested that microRNA-21 may become a novel potential biomarker for CRC. Consequently, this meta-analysis was designed to systematically review the values of microRNA-21 in CRC diagnosis.

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**Methodology:** Databases, including Cochrane library, PubMed, EMbase, Google Scholar, and Chinese National Knowledge Infrastructure, were scanned to retrieve relevant articles focusing on microRNA-21 in CRC diagnosis. Articles were then filtered according to the PRISMA statement and assessed by quality assessment of diagnosis accuracy studies-2. Sensitivity (SEN), specificity (SPE), positive likelihood ratio, negative likelihood ratio, and diagnostic odds ratio (DOR) were pooled using fixed-effects model or random-effects model. Summary receiver operating characteristic (SROC) curve and area under the curve (AUC) were used to estimate the overall diagnostic performance.

**Results:** A total of 15 studies, comprising 1268 CRC patients and 910 healthy controls, were enrolled in this meta-analysis. For serum miR-21, the pooled DOR, SEN, and SPE were 13.97 (95% CI: 8.44–23.11), 0.73 (95% CI: 0.69–0.77), and 0.83 (95% CI: 0.76–0.89), respectively; for plasma miR-21, the pooled DOR, SEN, and SPE were 8.03 (95% CI: 3.30–19.52), 0.67 (95% CI: 0.60–0.73), and 0.76 (95% CI: 0.69–0.81), respectively; and for fecal miR-21, the pooled DOR, SEN, and SPE were 7.06 (95% CI: 2.17–22.95), 0.33 (95% CI: 0.28–0.37), and 0.91 (95% CI: 0.88–0.93), respectively. Moreover, the AUC values of serum, plasma, and fecal miR-21 in CRC diagnosis were 0.8701, 0.8295, and 0.6742, respectively.

**Conclusion:** Blood miR-21 demonstrates good diagnostic performance, and serum samples are better than plasma samples in CRC diagnosis. For fecal miR-21, the sensitivity is unsatisfactory, but the specificity is favorable in predicting CRC patients.

*Keywords: MicroRNA-21; diagnosis; colorectal cancer; meta-analysis.*

# **1. INTRODUCTION**

In recent years, morbidity and mortality constitution of tumors published worldwide has demonstrated that [1-3] colorectal cancer is increasingly threatening to human health. Although early detection, early diagnosis, and early treatment are emphasized in clinical practice, limitations still exist in the present imaging technology for early diagnosis of colorectal cancer (CRC) [4,5]. For example, nuclear magnetic resonance imaging, computed tomography, and positron emission computed tomography show limitations of high cost, selective sizes of tumor tissues, and disconfirmation; air barium double-contrast radiography examination, transrectal ultrasound, and colonoscopy are usually rejected because of discomfort during operation [6]. Furthermore, serum tumor markers [7] and fecal occult-blood testing (FOBT) [8] exhibit low sensitivity and unsatisfactory specificity.

MicroRNAs (miRNAs) are evolutionarily conserved, endogenous, and non-coding RNA molecules, which consist of 17–25 nucleotides and can identify specific target mRNAs to act as negative gene regulators at post-transcriptional level. Considerable research has found that abnormal miRNA expression emerges in the occurrence and development of CRC, and tumorspecific miRNA exists in CRC patients' serum, plasma [9,10], feces [11] or tissue [12,13]; all of which can be stably detected and can predict tumor status; such discoveries ensure a promising application prospect for miRNAs as novel tumor biomarkers.

As a widely studied oncogenic miRNA, miR-21 has been notably upregulated in CRC patients. Previous studies proposed that miR-21 is positively correlated with pathological staging of CRC and negatively correlated with disease-free interval [14,15]. Additionally, miR-21 is considered a powerful prognostic marker applicable to CRC patients from various racial populations, and it can identify the progression of high-risk diseases in the early stage of cancer [16,17]. This meta-analysis was conducted to evaluate the clinical values of miR-21 in tissues, feces, serum, and plasma for systematic diagnosis of CRC.

# **2. MATERIALS AND METHODS**

# **2.1 Search Strategy**

All relevant articles were searched via the following electronic databases: Cochrane library, PubMed, EMbase, Google Scholar, and Chinese National Knowledge Infrastructure until January 10, 2015. Brief search strategies were as follows: ("Subject headings" or colorectal cancer or colon cancer or rectal cancer), ("Subject headings" or microRNA-21 or miRNA-21), ("Subject headings" or sensitivity or specificity), and other precise search strategies of PubMed (Table 1).

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#### **2.2 Inclusion and Exclusion Criteria**

#### **2.2.1 Inclusion criteria**

(1) all CRC patients were pathologically verified (gold standard); (2) CRC patients did not receive radiotherapy, chemotherapy, surgery nor any other treatment before sample collection; (3) studies had to contain evaluating indicators of microRNA-21 used alone for CRC diagnosis: sensitivity or specificity; (4) normal controls were set; (5) for similar literature published by the same author or research center, those with more recent publication time or higher influence factor were included, and resemble studies focusing on different indicators should all be involved; (6) cases of each group should be ≥10; (7) no restriction in age, gender, nationality, and race, and original articles published in either English or Chinese.

# **2.2.2 Exclusion criteria**

(1) duplicate publications; (2) missing data cannot be quantitatively synthesized; (3) nonhuman studies, cell researches, reviews, metaanalysis, meeting abstracts, or case reports.

# **2.3 Data Extraction and Quality Assessment**

On the basis of the above mentioned inclusion and exclusion criteria, two investigators (Chaohui ZHEN and Guojun YAO) independently<br>conducted literature selection, quality literature selection, quality<br>and data extraction. Any assessment, and data extraction. disagreement would be fully discussed between the former two researchers, or under the assistance of the third senior investigator to reach a consensus. The final enrolled articles were assessed by referring to quality assessment of diagnosis accuracy studies-2 (QUADAS-2) [18]. In diagnostic tests, QUADAS-2 was generally recognized as a quality assessing tool comprising four domains: (1) patient selection; (2) index test; (3) reference standard; and (4) flow and timing. The signaling questions of each domain are rated as "high," "unclear," and "low" to judge on risk of bias and applicability.



#### **Table 1. Search strategy in PubMed**

All statistical analyses were performed by Meta-DiSc and STATA12.0 statistical software. By extracting all relevant data (true positives, false positives, true negatives, and false negatives) from the enrolled studies, the pooled sensitivity (SEN), specificity (SPE), positive likelihood ratio (PLR), negative likelihood ratio (NLR), diagnostic odds ratio (DOR), and 95% confidence interval (95% CI) were obtained. Simultaneously, the investigators generated the summary receiver operating characteristic (SROC) curve and calculated the area under the curve (AUC). The analysis steps were as follows: (1) Heterogeneity test: a. The threshold effect was assessed using Spearman correlation coefficient [19]; a *P* value < 0.05 suggested the presence of the threshold effect requiring no meta-analysis; b. The nonthreshold effect was detected by Cochran-Q method and test of inconsistency index (*I*2) [20]; a low *P* value of <0.05 or a high *I*2 value of >50% suggested the existence of non-threshold effect, and meta-regression analysis was consequently used to determine the sources; (2) Metaregression analysis: The research-related covariates were compared, which were eliminated one-by-one based on the *P* values (from high to low); the covariates causing heterogeneity were determined, and subgroup analysis was performed; (3) Quantitative synthesis: a. Fixed-effects model (Mantel-Haenszel method) was used to combine the data if heterogeneity did not exist; b. Subgroup analysis or random-effects model (DerSimonian-Laird method) was applied to combine the data if non-threshold effect existed; (4) Publication bias was assessed by Begg's test and Egger's test [21]; a *P* value  $\geq$  0.05 indicated no publication bias.

# **3. RESULTS**

# **3.1 Characteristics of Eligible Studies**

After selection based on PRISMA statement [22], about 15 studies with a total of 1268 CRC patients and 910 healthy controls were eventually eligible. Fig. 1 shows the study selection process, and Table 2 lists the main characteristics of the eligible studies.

# **3.2 Quality Assessment**

The quality of the included studies was assessed using QUADAS-2 quality assessment. All 15 inclusions reached middle to high quality (Figs. 2 and 3). However, no blinding method was set in the detection of gold standard. Hence, a major bias was concentrated upon "index test"; bias also existed in "patient selection" because most of the research did not report nor recruit patients randomly.

# **3.3 Heterogeneity Test**

The heterogeneity among the studies is a critical factor influencing the accuracy of the synthesis. We first evaluated the existence of the threshold effect by calculating the Spearman correlation coefficient and *P* values (less than three studies about tissue samples were included; thus, heterogeneity test was not performed). The results showed that the SROC curves of blood (serum and plasma), serum, plasma, and fecal samples were not shoulder-shaped. For blood samples, the Spearman correlation coefficient and *P* value were −0.117 and 0.765, whereas 0.400 and 0.600 for serum samples, respectively; for plasma samples and fecal samples, the corresponding values were −0.500 and 0.391; and 0.500 and 0.667, respectively; all of which indicated that the threshold effect did not exist. Moreover, the *P* value of Cochran-Q method and the  $l^2$  value were employed to detect the existence of the non-threshold effect. The pooled SEN, SPE, PLR, NLR, and DOR of blood, plasma, and fecal samples obtained a *P* value of  $\leq$  0.05 and an  $l^2$  value of  $\geq$  50%, thus indicating that heterogeneity caused by non-threshold effect exists in these samples, and metaregression analysis should be performed. For serum samples, the pooled SPE, PLR, and DOR showed a  $P$  value of  $>0.05$  and an  $I^2$  value of <50%, which can be directly merged by FEM; the pooled SEN and NLR indicated a *P* value of  $\leq$  0.05 and an  $l^2$  value of  $\geq$  50%. Thus, metaregression was applied.

# **3.4 Meta-regression**

Meta-regression was employed to explain the research-related covariates causing nonthreshold effect. These covariates included race, internal control, assay type and study quality. Unfortunately, no satisfactory clues were found.

# **3.5 Quantitative Synthesis**

Research without non-threshold effect should use FEM to merge data. The potential heterogeneity caused by the non-threshold effect in studies of some sample types should also apply REM to merge data. For miR-21 in CRC diagnosis, the pooled SEN, SPE, PLR, and NLR of blood samples were 0.71 (95% CI: 0.67–0.74), 0.78 (95% CI: 0.74–0.83), 3.53 (95% CI: 2.35– 5.30), and 0.37 (95% CI: 0.27–0.49), respectively, and the pooled DOR and AUC were 10.57 (95% CI: 5.70–19.61) and 0.8447, respectively. Results of the subgroup analysis revealed that for serum samples, the pooled SEN, SPE, PLR, and NLR were 0.73 (95% CI: 0.69–0.77), 0.83 (95% CI: 0.76–0.89), 4.19 (95% CI: 2.91–6.04), and 0.30 (95% CI: 0.19–0.48), respectively, and the pooled DOR and AUC were 13.97 (95% CI: 8.44–23.11) and 0.8701, respectively; for plasma samples, the pooled SEN, SPE, PLR, and NLR were 0.67 (95% CI: 0.60–0.73), 0.76 (95% CI: 0.69–0.81), 3.11 (95% CI: 1.78–5.44), and 0.43 (95% CI: 0.29–0.64), respectively, the pooled DOR and AUC were 8.03 (95% CI: 3.30–19.52) and 0.8295, respectively; for fecal samples, the pooled SEN, SPE, PLR, and NLR were 0.33 (95% CI: 0.28–0.37), 0.91 (95% CI: 0.88–0.93), 4.77 (95% CI: 1.64–13.87), and 0.69 (95% CI: 0.51–0.93), respectively, the pooled DOR and AUC were 7.06 (95% CI: 2.17–22.95) and 0.6742. Generally, blood samples are more valuable than fecal samples for miR-21 in CRC diagnosis, and serum samples are more accurate than plasma. Table 3 shows the summary estimates of the

diagnostic criteria, and Figs. 4–8 illustrate the detailed data of blood, serum, plasma and feces.

#### **3.6 Publication bias**

Publication bias is another crucial factor influencing the estimating accuracy in metaanalysis of diagnostic tests. Begg's test and Egger's test were used in this meta-analysis. The *P* values for Begg's test and Egger's test were 0.404 and 0.187, respectively, which suggest that no publication bias existed among the included studies focusing on blood samples (Fig. 9). The number of studies on fecal samples was very small for use in Begg's test and Egger's test.

## **4. DISCUSSION**

To explore novel tumor markers for clinical applications, we performed this meta-analysis to review the values of miR-21 in different samples for systematic diagnosis of CRC. In this metaanalysis, discrepant expression levels of miR-21 in the blood, feces, and tissues showed obvious statistical significance between the CRC patients and the control individuals. For miR-21 in blood samples, the pooled SEN, SPE, PLR,



**Fig. 1. The flow chart of data identification and selection**

and NLR were 0.71 (95% CI: 0.67–0.74), 0.78 (95% CI: 0.74–0.83), 3.53 (95% CI: 2.35–5.30), and 0.37 (95% CI: 0.27–0.49), respectively, thus indicating prominent advantage in sensitivity and specificity of miR-21 in blood samples for the diagnosis of CRC. DOR [38] reflects the correlation between the diagnostic results and diseases. When DOR is >1, a larger value suggests better diagnostic performance. AUC [39] is used to evaluate the overall performance, and its optimal value tends to be 1. The pooled DOR and AUC of miR-21 in blood samples were 10.57 (95% CI: 5.70–19.61) and 0.8447, respectively, hence suggesting excellent performance of miR-21 in the diagnosis of CRC. In subgroup analyses, the pooled AUC, DOR, SEN, SPE, PLR, and NLR of serum samples were 0.8701, 13.97 (95% CI: 8.44–23.11), 0.73 (95% CI: 0.69–0.77), 0.83 (95% CI: 0.76–0.89), 4.19 (95% CI: 2.91–6.04), and 0.30 (95% CI: 0.19–0.48), respectively; for plasma samples, the pooled AUC, DOR, SEN, SPE, PLR, and NLR were 0.8295, 8.03 (95% CI: 3.30–19.52), 0.67 (95% CI: 0.60–0.73), 0.76 (95% CI: 0.69–0.81), 3.11 (95% CI: 1.78–5.44), and 0.43 (95% CI: 0.29–0.64), respectively. Considering the above evidence, the SEN, SPE, and overall performance of serum samples are better than that of the plasma samples, hence suggesting that anticoagulants may affect the extraction and detection of miRNAs. For fecal miR-21, the pooled AUC, DOR, SEN, SPE, PLR, and NLR were 0.6742, 7.06 (95% CI: 2.17–22.95), 0.33 (95% CI: 0.28–0.37), 0.91 (95% CI: 0.88–0.93), 4.77 (95% CI: 1.64–13.87), and 0.69 (95% CI: 0.51–0.93), respectively, indicating that fecal miR-21 exhibits favorable SPE, even though its SEN is unsatisfactory. When combined with multiple miRNAs, fecal miR-21 can decline the misdiagnosis rate of CRC. There were only 2 researches focusing on tissue miR-21 in the present study, and meta analysis was not recommended because the result of the combined data is not persuasive. Nevertheless, both single-center studies suggested high expression of miR-21 in colorectal cancer tissue, which will provide theoretical basis for future researches on other specimen types.



#### **Fig. 2. Quality assessment by QUADAS-2: summary of risk of bias and applicability concern**







# **Table 2. Main characteristics of included studies in this meta-analysis**

**Table 3. Summary estimates of diagnostic criteria and the 95% confidence interval**



*a means no AUC; at least three included studies are needed to synthesize SROC curve; b includes serum and plasma samples*

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**Fig. 4. The forest plots show the pooled diagnosis index of blood miR-21 for the diagnosis of CRC. The point efficiencies from each study are shown as squares, and the pooled efficiencies are shown as diamond. (A) Sensitivity and specificity; (B) PLR and NLR; (C) DOR and their 95% CI**



**Fig. 5. The forest plots show the pooled diagnosis index of serum miR-21 for the diagnosis of CRC. (A) Sensitivity and specificity; (B) PLR and NLR; (C) DOR and their 95% CI**

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**Fig. 6. The forest plots show the pooled diagnosis index of plasma miR-21 for the diagnosis of CRC. (A) Sensitivity and specificity; (B) PLR and NLR; (C) DOR and their 95% CI**



**Fig. 7. The forest plots show the pooled diagnosis index of feces miR-21 for the diagnosis of CRC. (A) Sensitivity and specificity; (B) PLR and NLR; (C) DOR and their 95% CI**



**Fig. 8. Summary receiver operating characteristic curves (SROC) of miR-21 describes the diagnostic performance. (A) The SROC curve of blood is symmetric, and the AUC is 0.8447. (B) The SROC curve of serum is symmetric, and the AUC is 0.8701. (C) The SROC curve of plasma is symmetric, and the AUC is 0.8295. (D) The SROC curve of feces is symmetric, and the AUC is 0.6742**



**Fig. 9. Publication bias from Begg's test and Egger's test are shown by funnel plots. The line is the regression line, and every point represents one study. No publication bias is shown to exist**

Some meta-analysis or systemic review articles are published in recent years on the diagnostic value of miR-21 in CRC [27,31]. All in all, the results indicated that miR-21 has a potential diagnostic value with moderate sensitivity and good specificity for CRC. Combined with our research, the diagnostic performance of miR-21 in blood samples is better than that of fecal samples, and serum samples are more beneficial than plasma for miRNA extraction and detection.

For any meta-analysis, completely avoiding the heterogeneity existing in the eligible studies [40] is indispensable; even publishers themselves prefer positive results, and publication bias thus emerges [41]. Fortunately, no threshold effect or publication bias was observed in our metaanalysis. Examination showed that non-threshold effect existed in the selected articles; however, meta-regression did not determine the responsible covariates. Hence, we combined the data by using REM to improve the accuracy of the combined results.

The prognosis of CRC patients is inversely related to tumor staging, early diagnosis, and early treatment, which can significantly decrease its mortality and recurrence. Undoubtedly, colonoscopy is a method with the maximum diagnostic performance, but it is rejected by most of patients because of its invasive nature [6]. Moreover, noninvasive examinations, including carcinoembryonic antigen [7] and FOBT [8] cannot work efficiently because of poor sensitivity and specificity. The occurrence, staging, metastasis, and recurrence of CRC are closely related to the abnormal expression of miRNAs, and their stable existence in serum, plasma [9,10], and feces [11] has facilitated clinical applications of miRNAs. Furthermore, our meta-analysis demonstrated that miR-21 in the blood yields preferable performance in the diagnosis of CRC; fecal miR-21 exhibits satisfactory specificity, and it can reduce the misdiagnosis rate in cooperation with other indicators. This evidence suggested that miR-21 is a potential powerful biomarker in CRC diagnosis.

Although its results are promising, this metaanalysis still has limitations. First, miR-21 has only recently emerged as a novel marker in CRC diagnosis, thus, our meta-analysis only includes a small sample size. Therefore, further validations of miR-21 in large cohort and independent studies are needed. Second, many other cancer types dysregulate miR-21. However, studies that distinguish CRC from other cancer types are lacking, which may limit the application of miR-21 in clinical settings. Further comprehensive studies are needed to solve this problem. Furthermore, the detecting methods for miRNAs are all based on qRT-PCR. Unified primers and reference miRNAs for qPCR analysis remain unavailable to date.

# **5. CONCLUSION**

In conclusion, our research demonstrated that miR-21 in blood samples has better diagnostic performance than that in feces, and serum samples are even better for the detection of miR-21. Fecal miR-21 exhibits favorable specificity but with poor sensitivity. However, several limitations exist in this meta-analysis. First, the implementation of the gold standard did not meet the requirements of the blind method; second, case selection did not strictly abide the continuous random inclusion; finally, small sample size was used in some selected studies. Consequently, blind-designed random studies in large scale should be strictly conducted by multicenter medical institutions locally and internationally to achieve more authoritative results.

# **CONSENT**

It is not applicable.

# **ETHICAL APPROVAL**

It is not applicable.

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# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# **REFERENCES**

1. He J, Zhao P, Chen WQ. Chinese cancer registry annual report. Military Medical Science Press, Beijing; 2012.

- 2. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. 2011; 61(2):69-90.
- 3. Siegel R, Desantis C, Jemal A. Colorectal cancer statistics. CA Cancer J Clin. 2014;64(2):104-17.
- 4. Liu NR, Wang Y, Liao FD, Huang SH, Chen R. Progress in early diagnosis of colorectal cancer. Acta Laser Biology Sinica. 2013;03.
- 5. Liang XM, Cheng YS. Progress in imaging diagnosis of colorectal cancer. World Chinese Journal of Digestology. 2008;11.
- 6. Walsh JM, Terdiman JP. Colorectal cancer screening: Scientific review. JAMA. 2003; 289(10):1288-96.
- 7. Eremina E. Use of serum tumor markers in the diagnosis of malignant tumors of the digestive system. Eksp Klin Gastroenterol. 2011;(7):91-8.
- 8. Collins JF, Lieberman DA, Durbin TE, Weiss DG. Accuracy of screening for fecal occult blood on a single stool sample obtained by digital rectal examination: A comparison with recommended sampling practice. Ann Intern Med. 2005;142(2):81-5.
- 9. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable bloodbased markers for cancer detection. Proc Natl Acad Sci U S A. 2008;105(30):10513- 8.
- 10. Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K, et al. Characterization of microRNAs in serum: A novel class of biomarkers for diagnosis of cancer and other diseases. Cell Res. 2008;18(10):997-1006.
- 11. Wu WK, Law PT, Lee CW, Cho CH, Fan D, Wu K, et al. MicroRNA in colorectal cancer: From benchtop to bedside. Carcinogenesis. 2011;32(3):247-53.
- 12. Bandres E, Cubedo E, Agirre X, Malumbres R, Zarate R, Ramirez N, et al. Identification by Real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. Mol Cancer. 2006;5:29.
- 13. Slaby O, Svoboda M, Fabian P, Smerdova T, Knoflickova D, Bednarikova M, et al. Altered expression of miR-21, miR-31, miR-143 and miR-145 is related to clinicopathologic features of colorectal cancer. Oncology. 2007;72(5-6):397-402.
- 14. Kulda V, Pesta M, Topolcan O, Liska V, Treska V, Sutnar A, et al. Relevance of miR-21 and miR-143 expression in tissue

samples of colorectal carcinoma and its liver metastases. Cancer Genet Cytogenet. 2010;200(2):154-60.

- 15. Schetter AJ, Leung SY, Sohn JJ, Zanetti KA, Bowman ED, Yanaihara N, et al. MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. JAMA. 2008; 299(4):425-36.
- 16. Shibuya H, Iinuma H, Shimada R, Horiuchi A, Watanabe T. Clinicopathological and prognostic value of microRNA-21 and microRNA-155 in colorectal cancer. Oncology. 2010;79(3-4):313-20.
- 17. Nielsen BS, Jorgensen S, Fog JU, Sokilde R, Christensen IJ, Hansen U, et al. High levels of microRNA-21 in the stroma of colorectal cancers predict short diseasefree survival in stage II colon cancer patients. Clin Exp Metastasis. 2011; 28(1):27-38.
- 18. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: A revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med. 2011;155(8):529-36.
- 19. Zamora J, Abraira V, Muriel A, Khan K, Coomarasamy A. Meta-DiSc: A software for meta-analysis of test accuracy data. BMC Med Res Methodol. 2006;6:31.
- 20. Dinnes J, Deeks J, Kirby J, Roderick P. A methodological review of how heterogeneity has been examined in systematic reviews of diagnostic test accuracy. Health Technol Assess. 2005; 9(12):1-113,iii.
- 21. Sterne JA, Gavaghan D, Egger M. Publication and related bias in metaanalysis: Power of statistical tests and prevalence in the literature. J Clin Epidemiol. 2000;53(11):1119-29.
- 22. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. Int J Surg. 2010;8(5):336-41.
- 23. Ogata-Kawata H, Izumiya M, Kurioka D, Honma Y, Yamada Y, Furuta K, et al. Circulating exosomal microRNAs as biomarkers of colon cancer. PLoS One. 2014;9(4):e92921.

DOI: 10.1371/journal.pone.0092921 24. Toiyama Y, Takahashi M, Hur K, Nagasaka T, Tanaka K, Inoue Y, et al. Serum miR-21 as a diagnostic and prognostic biomarker in colorectal cancer. J Natl Cancer Inst. 2013;105(12):849-59.

- 25. Wang B, Zhang Q. The expression and clinical significance of circulating microRNA-21 in serum of five solid tumors. J Cancer Res Clin Oncol. 2012; 138(10):1659-66.
- 26. Liu GH, Zhou ZG, Chen R, Wang MJ, Zhou B, Li Y, et al. Serum miR-21 and miR-92a as biomarkers in the diagnosis and prognosis of colorectal cancer. Tumour Biol. 2013;34(4):2175-81.
- 27. Du M, Liu S, Gu D, Wang Q, Zhu L, Kang M, et al. Clinical potential role of circulating microRNAs in early diagnosis of colorectal cancer patients. Carcinogenesis. 2014; 35(12):2723-30.
- 28. Kanaan Z, Rai SN, Eichenberger MR, Roberts H, Keskey B, Pan J, et al. Plasma miR-21: A potential diagnostic marker of colorectal cancer. Ann Surg. 2012; 256(3):544-51.
- 29. Luo X, Stock C, Burwinkel B, Brenner H. Identification and evaluation of plasma microRNAs for early detection of colorectal cancer. PLoS One. 2013;8(5):e62880. DOI: 10.1371/journal.pone.0062880
- 30. Zanutto S, Pizzamiglio S, Ghilotti M, Bertan C, Ravagnani F, Perrone F, et al. Circulating miR-378 in plasma: A reliable, haemolysis-independent biomarker for colorectal cancer. Br J Cancer. 2014; 110(4):1001-7.
- 31. Zhang H, Li P, Ju H, Pesta M, Kulda V, Jin W, et al. Diagnostic and prognostic value of microRNA-21 in colorectal cancer: An original study and individual participant data meta-analysis. Cancer Epidemiol Biomarkers Prev. 2014;23(12):2783-92.
- 32. Kanaoka S, Kuriyama S, Iwaizumi M, Yamada T, Sugimoto M, Osawa S, et al. Potential usefulness of fecal immunochemical test plus fecal microrna assay as a marker for colorectal cancer screening. Gastroenterology. 2013;144(5).
- 33. Wu CW, Ng SS, Dong YJ, Ng SC, Leung WW, Lee CW, et al. Detection of miR-92a and miR-21 in stool samples as potential screening biomarkers for colorectal cancer and polyps. Gut. 2012;61(5):739-45.
- 34. Koga Y, Yasunaga M, Takahashi A, Kuroda J, Moriya Y, Akasu T, et al. MicroRNA expression profiling of exfoliated colonocytes isolated from feces for colorectal cancer screening. Cancer Prev Res (Phila). 2010;3(11):1435-42.
- 35. Kuriyama S, Hamaya Y, Yamada T, Sugimoto M, Osawa S, Sugimoto K, et al. Fecal microrna assays as a marker for<br>colorectal cancer screening. cancer screening. Gastroenterology. 2012;142(5).
- 36. Liu K, Li G, Fan C, Zhou X, Wu B, Li J, et al. Increased expression of microRNA-21 and its association with chemotherapeutic response in human colorectal cancer. J Int Med Res. 2011;39(6):2288-95.
- 37. Omrane I, Kourda N, Stambouli N, Privat M, Medimegh I, Arfaoui A, et al. MicroRNAs 146a and 147b biomarkers for colorectal tumor's localization. Biomed Res Int. 2014;2014:584852. DOI: 10.1155/2014/584852
- 38. Glas AS, Lijmer JG, Prins MH, Bonsel GJ, Bossuyt PM. The diagnostic odds ratio: A single indicator of test performance. J Clin Epidemiol. 2003;56(11):1129-35.
- 39. Jones CM, Athanasiou T. Summary receiver operating characteristic curve analysis techniques in the evaluation of diagnostic tests. Ann Thorac Surg. 2005;79(1):16-20.
- 40. Lijmer JG, Bossuyt PM, Heisterkamp SH. Exploring sources of heterogeneity in systematic reviews of diagnostic tests. Stat Med. 2002;21(11):1525-37.
- 41. Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for<br>publication bias. Biometrics. 1994: publication bias. Biometrics. 1994; 50(4):1088-101.

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