



## **Impact of Different Arbuscular Mycorrhizal Fungi on Nutrient Status and Their Uptake by *Melia azedarach* Seedling**

**Gulab<sup>a\*</sup>, Ramesh Verma<sup>b</sup>, K. S. Ahlawat<sup>a</sup>, Rakesh Chugh<sup>c</sup>, Chhavi Sirohi<sup>a</sup>,  
Satish Kumar Mehta<sup>d</sup>, Charan Singh<sup>e</sup> and Mamta Khaiper<sup>a</sup>**

<sup>a</sup> Department of Forestry, CCS Haryana Agricultural University, Hisar-125004, Haryana, India.

<sup>b</sup> Krishi Vigyan Kender, CCS Haryana Agricultural University, Kaithal-136027, Haryana, India.

<sup>c</sup> Department of Plant Pathology, CCS Haryana Agricultural University, Hisar-125004, Haryana, India.

<sup>d</sup> SNIATTE, Directorate of Extension Education, CCSHAU, Hisar-125004, India.

<sup>e</sup> Department of Soil Science, CCS Haryana Agricultural University, Hisar-125004, Haryana, India.

### **Authors' contributions**

*This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.*

### **Article Information**

DOI: 10.9734/IJPSS/2022/v34i1130941

### **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/85299>

**Original Research Article**

**Received 17 January 2022**

**Accepted 24 March 2022**

**Published 28 March 2022**

### **ABSTRACT**

*Melia azedarach* belongs to the family *Meliaceae*, is a deciduous tree with a short bole and a spreading crown and one of the most important trees in arid and semi-arid area. The arbuscular mycorrhizal fungi (AMF) are soil microorganisms composing the essential components of the sustainable soil-plant system. These AMF form extensive extraradical mycelia which increases accessible soil volume for the plant to absorb phosphorus and water. The effect of three different species of *Glomus* (*G. mosseae*, *G. intraradices* and *G. fasciculatum*) of AM fungi inoculated with *Melia azedarach* drupes was observed in the nursery of the Forestry Department, CCS Haryana Agricultural University, Hisar during 2019. The results revealed that application of three *Glomus* species in combination produced synergistic increase in soil macro and micro nutrient contents after harvest i.e. N = 119.86 kg ha<sup>-1</sup>, P = 16.91 kg ha<sup>-1</sup>, K = 260.86 kg ha<sup>-1</sup>, Fe = 5.95 ppm, Zn = 0.66 ppm, Cu = 0.42 ppm and Mn = 1.63 ppm. Similarly, there was significantly higher macro and micronutrient (N = 1.39 %, P = 0.27 %, K = 0.63 %, Fe = 90.43 ppm, Zn = 32.24 ppm, Cu = 9.96 ppm and Mn = 31.28 ppm) concentrations in leaves after 180 days in the treatments when soil was

\*Corresponding author: E-mail: [gulab.khatri.907@gmail.com](mailto:gulab.khatri.907@gmail.com);

inoculated with all three *Glomus* species. The seedlings uptake of macro and micronutrient was also found highest when three *Glomus* species were inoculated artificially and simultaneously in soil. Inoculation of the AM fungi improved soil fertility, nutrient content and uptake in *Melia azedarach*.

**Keywords:** AMF; *G. mosseae*; *G. intradices*; *G. fasciculatum*; macro and micro nutrient.

## 1. INTRODUCTION

India is the seventh-largest country globally, with a total area of 7, 67,419 square km under the tree. The country's total forest area is 24.62 per cent of its geographical area comprising 21.71 per cent of the forest cover and 2.91 per cent tree cover and most growing states are Madhya Pradesh, Arunachal Pradesh, Maharashtra, Orissa, Chhattisgarh and Haryana. In Haryana, forests cover shares only 3.63 per cent of the total geographical area [1]. *Melia azedarach* (family Meliaceae) is a moderate deciduous tree with a short bole and a spreading crown, bipinnate or tri-pinnate leaves and a dark grey bark with shallow longitudinal furrows [2]. This plant has a reasonably coarse root system with very few root hairs, which are common characteristics of plants that are responsive to arbuscular mycorrhizal symbiosis. The arbuscular mycorrhizal fungi (AMF) are soil microorganisms having extensive extraradical mycelia which increases phosphorus and water absorption [3]. These fungi provide their host with various benefits including higher phosphorus absorption (Gossus and Mohammad, 2009), defending root against diseases [4], plant growth-promoting hormone [5] and boosting plant growth and productivity [6]. The mycorrhizal fungi are naturally occurring species that form symbiotic relationships with almost all plants inside the roots. This also provides a perfect ecological niche that is necessary for fungal growth and development including the completion of the sexual cycle [7]. The arbuscules are branched hyphae, within the root cells and this is a connection for nutrient exchange between fungi and the host plant [8]. Fungus improves nutrition, growth, drought resistance and provides nutrients and water to the host plant [9]. Besides the direct effects of AM fungi on the cycling of nutrients and plant performance, there are several indirect effects of AMF for improving soil fertility via enmeshment of soil particles for better soil aggregation ([10] and nutrient maintenance [11]. Therefore, farmers' field management through a greater abundance of AM fungi could have improved nutrient accessibility and availability [12]. However, sustaining AM fungi

especially mycelia, needs a supply of plentiful carbohydrates from host plants [13]. In Haryana, agroforestry is distributed throughout arid and semi-arid regions [14], which could act as perennial AM fungi hosts and thus satisfy AM fungi continuous carbohydrate demand [15]. Some trees are exhaustive for nutrients but not all so, it depends on tree species for their effect on soil nutrient status, and differences in their relative availability which may further affect the AM fungi symbiosis relationship functioning [16] and AM fungi growth strategies [17]. Furthermore, it is vital to contemplate the overall effect of tree species with AM fungi inoculation on nutrients trade-off ultimately affecting soil fertility status, tree growth, and biomass production, including potential trade-offs regarding competition for light, water, and nutrients [18,19]. Globally, it remains an open question whether trees grown with AM fungi can rejuvenate and preserve soil fertility and, thus, boost crop performance. However, few studies have been made to isolate and select effective AM fungi for inoculation in *Melia azedarach* in nursery conditions. Keeping in view of all these factors the present study was planned to find out the effect of different AM fungi (*Glomus* sp.) on the growth and nutrient uptake of *Melia azedarach* seedling.

## 2. MATERIALS AND METHODS

The study was carried out in the nursery of Forestry Department, CCS Haryana Agricultural University, Hisar (20° 10' N lat., 75° 46' E long., alt. 215 m msl), situated in the semi-arid region of North-Western India. The soil types were Typic Ustochrept with sandy loam texture. The climate of Hisar (Haryana) is semi-arid with hot and dry desiccating winds accompanied by frequent dust storms with high velocity in summer months, severe cold during in winter months and humid warm during monsoon rainy season. The mean monthly maximum and minimum temperature sometimes exceed 48°C on hot summer days. The relative humidity varies from 5 to 100 per cent, while temperature below freezing point accompanied by frost in winter is usually experienced in this region. The

characteristics of mother plant i.e. plant height; DBH and clear bole height were 9.1 m, 0.4 m and 4.2 m, respectively. An average of 100 drupes were used for the experiment and traits such as fruit length (15 mm), fruit breadth (10 mm), fruit weight (0.32 g), fruit thickness (1.7 mm) and test weight (24 g) were recorded. The soil and rootlets from the root horizon of *Glomus mosseae*, *Glomus fasciculatum* and *Glomus intraradices* inoculated wheat plants were used to inoculate soil before sowing of drupes of *Melia azedarach* in seven treatments [20]. The experimental soil was collected from Balsamand Research Farm, CCS HAU, Hisar. The soil was drenched with formaldehyde, and immediately covered with an air-tight polythene sheet for 5 days. The drupes of *Melia azedarach* were collected from plus tree at Hisar and were sown at about 2-3 cm deep in polybags of one kg soil capacity inoculated with different *Glomus* species as per detail given below. This experiment was carried out with completely randomized design and each treatment having three replications.

There were seven treatments of soil inoculation of three different *Glomus* sp. inoculated singly as well as in combination before sowing and control (uninoculated AM fungi) as shown below:-

- T<sub>1</sub> - Treated soil +*G. mosseae* (450-500 sporocarps kg<sup>-1</sup> soil)
- T<sub>2</sub> - Treated soil +*G. intraradices* (450-500 sporocarps kg<sup>-1</sup> soil)
- T<sub>3</sub> - Treated soil +*G. fasciculatum* (450-500 sporocarps kg<sup>-1</sup> soil)
- T<sub>4</sub> - Treated soil + *G. mosseae* + *G. intraradices* (225-250 sporocarps of each AM fungi kg<sup>-1</sup> soil)
- T<sub>5</sub> - Treated soil +*G. mosseae* + *G. Fasciculatum* (225-250 sporocarps of each AM fungi kg<sup>-1</sup> soil)
- T<sub>6</sub> - Treated soil +*G. intraradices* + *G. Fasciculatum* (225-250 sporocarps of each AM fungi kg<sup>-1</sup> soil)
- T<sub>7</sub> - Treated soil +*G. mosseae* + *G. intraradices* + *G. fasciculatum* (150-165 sporocarps of each AM fungi kg<sup>-1</sup> soil)
- T<sub>8</sub> - Control (un-inoculated)

The soil samples were collected before and after harvesting of *M. azedarach* seedlings. These samples were air-dried and passed through a 2 mm sieve before determining macro and micronutrients [21]. The alkaline permanganate method described by Subbiah and Asija [22] was used to determine available nitrogen in soil. The available phosphorus was determined spectrophotometrically by Olsen's method [23]

and blue colour intensity was measured at 660 nm wavelength using a red filter. Neutral normal NH<sub>4</sub>OAC solution was used for estimation of available Potassium on flame photometer [24]. DTPA-TEA buffer (0.005 M DTPA + 0.001 M CaCl<sub>2</sub> + 0.1 M TEA, pH 7.3) as proposed by Lindsay and Norvell [25] was used for the determination of available micronutrients (Fe, Cu, Zn and Mn) concentration using atomic absorption spectrophotometer. The plant leaf samples were ground and digested with the di-acid mixture (nitric acid and perchloric acid). The colorimetric method proposed by Lindner [26] was used to estimate total nitrogen in the plant. Blue filter at 440 nm wavelength was used to read the intensity of orange colour on spectrophotometer. In the plant sample, total phosphorus was determined by Vanado-molybdophosphoric yellow colour method proposed by Koenig and Johnson [27]. The potassium was determined by a Flame photometer using the standard curve [28]. The micronutrient concentration in aliquot was determined spectrophotometrically at most sensitive wavelengths (Fe-248.7 nm, Zn-213.7 nm, Cu-324.6 nm and Mn-279.5 nm), Elwell and Gridley, [29].

**Macronutrient uptake (g plant<sup>-1</sup>) =**  

$$[\text{Macronutrient content in plant (\%)} \times \text{total dry weight of corresponding plant (g)}] / 100$$

**Micronutrient uptake (mg plant<sup>-1</sup>) =**  

$$[\text{Micronutrient content in plant (ppm)} \times \text{total dry weight of corresponding plant (g)}] / 10^3$$

Hyphae contribution (HC) was calculated using the following formula [30]:

$$\text{HC} = (\text{X uptake of the mycorrhizal plants} - \text{X uptake of the non-mycorrhizal plants}) / (\text{X uptake of the mycorrhizal plants} \times 100)$$

Where, X= respective nutrient (P/Zn)

### 3. RESULTS AND DISCUSSION

The data presented in Fig. 1 showed that available N, P and K contents were found maximum 119.86, 16.91 and 260.86 kg ha<sup>-1</sup>, respectively in treatment T<sub>7</sub> followed by treatment (T<sub>5</sub>) when *G. mosseae* and *G. fasciculatum* were applied in soil together i.e. 118.99, 15.81 and 258.87 kg ha<sup>-1</sup>, respectively after harvest of seedlings. However, the nutrient content of N and K were found non-significant, when drupes were inoculated individually with *G. mosseae*

(T<sub>1</sub>), *G. intraradices* (T<sub>2</sub>) and *G. fasciculatum* (T<sub>3</sub>). The phosphorus was also non-significant when drupes were inoculated singly with *G. mosseae* (T<sub>1</sub>) and *G. fasciculatum* (T<sub>3</sub>). The maximum Fe, Cu, Zn and Mn contents were recorded in T<sub>7</sub> (5.95, 0.42, 0.66 and 1.63 ppm, respectively) followed by treatment of *G. mosseae* + *G. fasciculatum* (5.66, 0.39, 0.63 and 1.58 ppm, respectively) and minimum in control (un-inoculated). The iron (Fe) and manganese (Mn) were found non-significant after harvest in soils treated with *G. mosseae* (T<sub>1</sub>), *G. intraradices* (T<sub>2</sub>) and *G. fasciculatum* (T<sub>3</sub>) whereas, *G. mosseae* and *G. fasciculatum* treatment were found non-significant for Cu and Zn content in the soil after harvest of *Melia azedarach* seedlings (Fig. 2). There was significantly higher N, P and K (%) content in six months old seedlings in T<sub>7</sub> (1.39, 0.27 and 0.63 %, respectively) and minimum in control (T<sub>8</sub>) with 1.27, 0.10 and 0.51 %, respectively (Table 1). However, the nutrient content of N and K (%) was found non-significant in seedlings after harvest in treatments of *G. mosseae*, *G. intraradices* and *G. fasciculatum* whereas, *G.*

*mosseae* and *G. fasciculatum* treatments were non-significant with each other for P (%) content. Among all the treatments, Fe, Cu, Zn and Mn (ppm) contents were recorded maximum in T<sub>7</sub> (90.43, 9.96, 32.24 and 31.28 ppm, respectively) whereas, minimum in control (77.79, 5.58, 15.45 and 25.12 ppm, respectively). However, in treatments with *G. mosseae*, *G. intraradices* and *G. fasciculatum* the content of Fe and Mn (85.56, 85.23, 85.43 ppm and 27.61, 27.45, 27.52 ppm, respectively) were found non-significant in seedlings after harvest, whereas, Cu and Zn content (7.15, 7.11 ppm and 21.87, 21.71 ppm, respectively) were found non-significant in seedlings of *M. azedarach* after harvest in *G. mosseae* and *G. fasciculatum* treatment. Basumatary et al. [31] reported a significant higher nutrient concentration of N, P, K and Carbon in soil when the soil was infested with *Acaulospora* sp. and *Glomous* sp. over control. The mycorrhizae play an important role and helps in nutrient cycling to establish plants [32]. It helps in better availability of nutrients to the plants mainly N and K with AM fungi inoculation in star fruits [33].

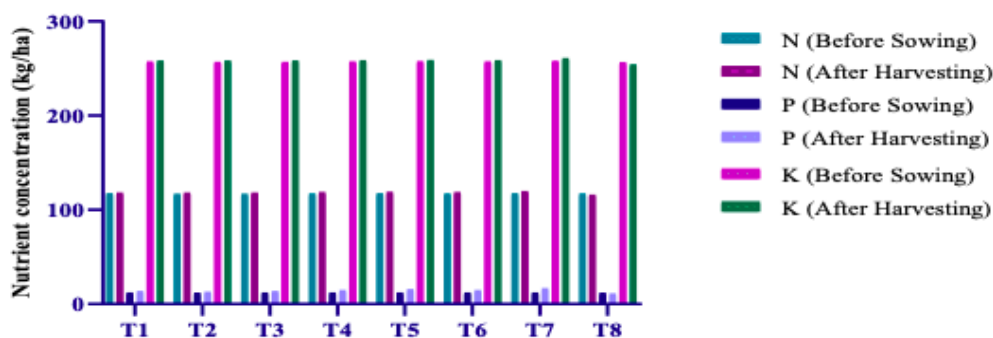


Fig. 1. Available nitrogen, phosphorus and potassium in soil before sowing and after harvest of *M. azedarach* seedlings

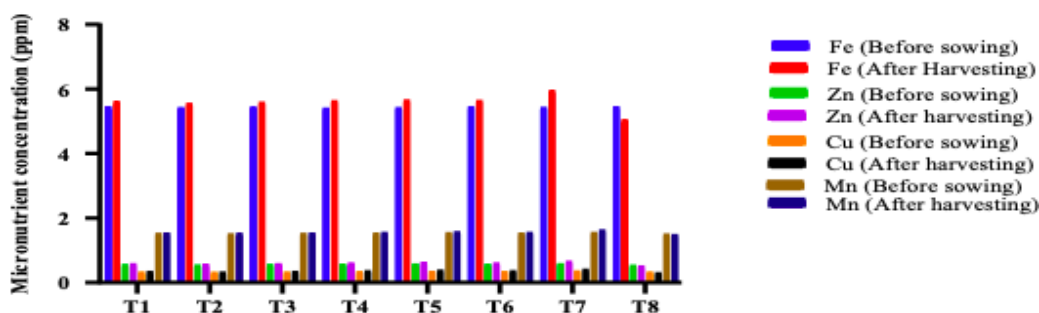


Fig. 2. Soil DTPA extractable Fe, Zn, Cu and Mn content before sowing and after harvest of *Melia azedarach* seedlings

**Table 1. Macro and micronutrient content in *Melia azedarach* seedlings after 180 days of sowing**

Treatments		Nutrients content						
		N (%)	P (%)	K (%)	Fe (ppm)	Cu (ppm)	Zn (ppm)	Mn (ppm)
T <sub>1</sub>	<i>Glomus mosseae</i>	1.31	0.17	0.55	85.56	7.15	21.87	27.61
T <sub>2</sub>	<i>G. intraradices</i>	1.30	0.14	0.54	85.23	6.45	18.67	27.45
T <sub>3</sub>	<i>G. fasciculatum</i>	1.31	0.17	0.54	85.43	7.11	21.71	27.52
T <sub>4</sub>	<i>G. mosseae</i> + <i>G. intraradices</i>	1.34	0.20	0.58	86.81	7.82	24.97	28.83
T <sub>5</sub>	<i>G. mosseae</i> + <i>G. fasciculatum</i>	1.35	0.23	0.59	87.02	8.77	27.91	28.99
T <sub>6</sub>	<i>G. intraradices</i> + <i>G. fasciculatum</i>	1.34	0.20	0.58	86.27	7.78	24.91	28.76
T <sub>7</sub>	<i>G. mosseae</i> + <i>G. intraradices</i> + <i>G. fasciculatum</i>	1.39	0.27	0.63	90.43	9.96	32.24	31.28
T <sub>8</sub>	Control (un-inoculated)	1.27	0.10	0.51	77.79	5.58	15.45	25.12
CD at 5%		0.02	0.02	0.02	2.13	0.46	2.25	1.45

**Table 2. Macro and micronutrient uptake in *Melia azedarach* after 180 days of sowing**

Treatments		Nutrients uptake						
		g/plant			mg/plant			
		N	P	K	Fe	Cu	Zn	Mn
T <sub>1</sub>	<i>G. mosseae</i>	0.26	0.04	0.10	1.70	0.14	0.43	0.55
T <sub>2</sub>	<i>G. intraradices</i>	0.23	0.02	0.09	1.52	0.12	0.33	0.49
T <sub>3</sub>	<i>G. fasciculatum</i>	0.25	0.04	0.10	1.64	0.14	0.42	0.54
T <sub>4</sub>	<i>G. mosseae</i> + <i>G. intraradices</i>	0.30	0.06	0.13	1.91	0.17	0.55	0.60
T <sub>5</sub>	<i>G. mosseae</i> + <i>G. fasciculatum</i>	0.33	0.08	0.16	2.01	0.22	0.69	0.67
T <sub>6</sub>	<i>G. intraradices</i> + <i>G. fasciculatum</i>	0.29	0.06	0.13	1.88	0.17	0.54	0.63
T <sub>7</sub>	<i>G. mosseae</i> + <i>G. intraradices</i> + <i>G. fasciculatum</i>	0.37	0.11	0.19	2.41	0.27	0.86	0.83
T <sub>8</sub>	Control (un-inoculated)	0.19	0.01	0.08	1.22	0.09	0.24	0.39
CD at 5%		0.03	0.01	0.02	0.21	0.01	0.05	0.07

**Table 3. Hyphal contribution (%) of AM fungi for Phosphorus and Zinc**

Treatments		180 DAS	
		HC (%) for P	HC (%) for Zn
T <sub>1</sub>	<i>G. mosseae</i>	75.0	44.18
T <sub>2</sub>	<i>G. intraradices</i>	50.0	27.27
T <sub>3</sub>	<i>G. fasciculatum</i>	70.0	42.85
T <sub>4</sub>	<i>G. mosseae</i> + <i>G. intraradices</i>	83.3	56.36
T <sub>5</sub>	<i>G. mosseae</i> + <i>G. fasciculatum</i>	87.5	65.21
T <sub>6</sub>	<i>G. intraradices</i> + <i>G. fasciculatum</i>	83.3	55.55
T <sub>7</sub>	<i>G. mosseae</i> + <i>G. intraradices</i> + <i>G. fasciculatum</i>	90.9	72.09
T <sub>8</sub>	Control (un-inoculated)	00.0	0

A significantly higher N, P and K uptake was recorded in six months old seedlings in T<sub>7</sub> (0.370, 0.11 and 0.19 (g plant<sup>-1</sup>), respectively and minimum in control with 0.19, 0.01 and 0.08 (g plant<sup>-1</sup>), respectively (Table 2). Caglar and Akgun [34] reported that uptake of nitrogen in the leaf of

*Pistacia* seedlings was enhanced upon inoculation with *Glomus* species. Among all the treatments Fe, Cu, Zn and Mn uptake were recorded maximum in interactive treatments of all species (T<sub>7</sub>) (2.41, 0.27, 0.86 and 0.83 mg plant<sup>-1</sup>, respectively) whereas, minimum was recorded

in control (1.22, 0.09, 0.24 and 0.39 mg plant<sup>-1</sup>, respectively). These result are in corroboration with the findings of many researchers viz., Jamaluddin and Shukla [35], Muthukumar et al. [36], Mohan and Sandeep [37] and Ortiz et al. [38] who reported an increase in nutrient uptake in different plant species with the application of different AM fungi species. The maximum Hyphal contribution (%) for P and Zn was found in T<sub>7</sub> followed by T<sub>5</sub> while under control there was no HC recorded (Table 3).

#### 4. CONCLUSION

The drupes of *M. azedarach* treated concomitantly with *G. mosseae*, *G. intraradices*, *G. fasciculatum* enhanced the soil nutrients especially P and Zn. The nutrients content in *M. azedarach* seedlings such as N (1.39 %), P (0.27 %), K (0.63 %), Fe (90.43 ppm), Cu (9.96 ppm), Zn (32.24 ppm) and Mn (31.28 ppm) were also found significantly higher in drupes sown in soil concomitantly treated with *G. mosseae*, *G. intraradices* and *G. fasciculatum*. The macro and micronutrient uptake was also found significantly higher in drupes sown in soil treated with all three *Glomus* species as cited above. Thus, it is concluded that the concomitant application of all three *Glomus* species have the potential to improve soil fertility, nutrient content and uptake of *M. azedarach*.

#### ACKNOWLEDGEMENT

Authors are sincerely thankful to the Department of Forestry, Soil Science and Plant Pathology, CCS HAU, Hisar for their kind support in carrying out this experiment.

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. India State of Forest Report. Forest Survey of India, Ministry of Environment and Forests, released; 2021.
2. Troup RS. The silviculture of Indian Trees. Clarendon Press, Oxford. 1921;1195.
3. Dierks J, Blaser-Hart WJ, Gamper HA, Nyoka IB, Barrios E, Six J. Trees enhance abundance of arbuscular mycorrhizal fungi, soil structure, and nutrient retention in low-input maize cropping systems. *Agriculture, Ecosystems and Environment*. 2021;318:107487.
4. Bakhtiar Y, Yahya S, Sumaryono W, Sinaga MS, Budi SW, Tajuddin T. Isolation and identification of Mycorrhizosphere bacteria and their antagonistic effects towards *Ganoderma boninense in vitro*. *Microbiology Indonesia*. 2010;4(2):9.
5. Herrera-Medina MJ, Steinkellner S, Vierheilig H, Bote JAO, Garrido JMG. Abscisic acid determines arbuscule development and functionality in the tomato arbuscular mycorrhiza. *New Phytologist*. 2007;175(3):554-564.
6. Duponnoisa R, Colombeta A, Hienb V, Thioulouse J. The mycorrhizal fungus *Glomus intraradices* and rock phosphate amendment influence plant growth and microbial activity in the rhizosphere of *Acacia holosericea*. *Soil Biochemistry*. 2005;37(8):1460-1468.
7. Sandeep C, Mohan V, Viswanath S. Significance of ectomycorrhizae in forest ecosystems of India. *International Journal of Plant, Animal and Environment Sciences*. 2015;5:23-31.
8. Afzal M, Yousaf S, Reichenauer TG, Kuffner M, Sessitsch A. Soil type affects plant colonization, activity and catabolic gene expression of inoculated bacterial strains during phytoremediation of diesel. *Journal of Hazardous Materials*. 2011;186(2):1568-1575.
9. Luyindula N, Haque I. Effect of rhizobium inoculation and phosphorus on growth and nitrogen fixation in tree grown on vertisols. In K. Mulongoy, M. Gueye and D.S.F. Spencer. *Biological nitrogen fixation and sustainability of tropical Agriculture*, New York. 1992;109 -113.
10. Rillig MC, Aguilar-Trigueros CA, Bergmann J, Verbruggen E, Veresoglou SD, Lehmann A. Plant root and mycorrhizal fungal traits for understanding soil aggregation. *New Phytologist*. 2015;205(4):1385-1388.
11. Cavagnaro TR, Bender SF, Asghari HR, Van der Heijden MG. The role of arbuscular mycorrhizas in reducing soil nutrient loss. *Trends in Plant Science*. 2015;20(5):283-290.
12. Sato T, Ezawa T, Cheng W, Tawaraya K. Release of acid phosphatase from extraradical hyphae of arbuscular mycorrhizal fungus *Rhizophagu sclarus*. *Soil Science and Plant Nutrition*. 2015;61(2):269-274.
13. Raven JA, Lambers H, Smith SE, Westoby M. Costs of acquiring phosphorus by

- vascular land plants: patterns and implications for plant coexistence. *New Phytologist*. 2018;217(4):1420-1427.
14. Handa AK, Sirohi C, Chavan SB, Dhillon RS, Ahlawat KS, Rizvi RH. Agroforestry in Haryana: Status and way forward. *Indian Journal of Agroforestry*. 2020;22(1):1-10.
  15. Bainard LD, Klironomos JN, Gordon AM. Arbuscular mycorrhizal fungi in tree-based intercropping systems: a review of their abundance and diversity. *Pedobiologia*. 2011;54(2):57-61.
  16. Johnson NC. Resource stoichiometry elucidates the structure and function of arbuscular mycorrhizas across scales. *New Phytologist*. 2010;185(3):631-647.
  17. Chagnon PL, Bradley RL, Maherali H, Klironomos JN. A trait-based framework to understand life history of mycorrhizal fungi. *Trends in Plant Science*. 2013;18(9):484-491.
  18. Wartenberg AC, Blaser WJ, Gattinger A, Roshetko JM, Van Noordwijk M, Six J. Does shade tree diversity increase soil fertility in cocoa plantations?. *Agriculture, Ecosystems and Environment*. 2017;248:190-199.
  19. Blaser WJ, Opong J, Hart SP, Landolt J, Yeboah E, Six J. Climate-smart sustainable agriculture in low-to-intermediate shade agroforests. *Nature Sustainability*. 2018;1(5):234-239.
  20. Spagnoletti FN, Cornero M, Chiochio V, Lavado RS, Roberts IN. *Arbuscular mycorrhiza* protects soybean plants against *Macrophomina phaseolina* even under nitrogen fertilization. *European Journal of Plant Pathology*. 2020;156(3):839-849.
  21. Antil RS, Singh A, Dahiya SS. Practical manual for soil and plant analysis. Department of Soil Science, CCS Haryana Agricultural University, Hisar-125004; 2002.
  22. Subbiah BV, Asija GL. A rapid procedure for assessment of available nitrogen in soils. *Current Science*. 1956;25:259-260.
  23. Olsen SR. Estimation of available phosphorus in soils by extraction with sodium bicarbonate. United States Department of Agriculture; Washington. 1954;2:25-43.
  24. Hanway JJ, Heidel H. Soil analysis methods as used in Iowa state college soil testing laboratory. *Iowa Agriculture*. 1952;57:1-31.
  25. Lindsay WL, Norvell WA. Development of a DTPA soil test for zinc, iron, manganese and copper. *Soil Science Society of America Journal*. 1978;42(3):421-428.
  26. Lindner RC. Rapid analytical methods for some of the more common inorganic constituents of plant tissues. *Plant Physiology*. 1944;19(1):76.
  27. Koenig R, Johnson C. Colorimetric determination of phosphorus in biological materials. *Industrial and Engineering Chemistry Analytical Edition*. 1942;14(2):155-156.
  28. Jackson ML. *Soil chemical analysis*. Prentice Hall of India. 1973;498.
  29. Elwell WT, Gridley JAF. *Atomic absorption spectrophotometer* ypergamon press Ltd. London, W-1; 1967.
  30. Kothari SK, Marschner H, Römheld V. Contribution of the VA mycorrhizal hyphae in acquisition of phosphorus and zinc by maize grown in a calcareous soil. *Plant and Soil*. 1991;131(2):177-185.
  31. Basumatary N, Parkash V, Tamuli AK, Saikia AJ, Teron R. *Arbuscular mycorrhizal* inoculation affects growth and rhizospheric nutrient availability in *Hevea brasiliensis* (Willd. ex A. Juss.) Mull. Arg. clones. *International Journal of Current Biotechnology*. 2014;2(7):14-21.
  32. Dhar PP, Mridha MAU. Arbuscular mycorrhizal associations in different forest tree species of Hazarikhil forest of Chittagong, Bangladesh. *Journal of Forestry Research*. 2012;23(1):115-122.
  33. Filho JAV, Mendonça Freitas MS, Martins MA, dos Santos PC, Cordeiro de Carvalho AJ. Arbuscular mycorrhizal fungi and phosphate fertilization on star fruit tree seedlings. *Revista Brasileira de Ciências Agrárias*. 2017;12(1):14-19.
  34. Caglar S, Akgun A. Effects of vesicular-arbuscular mycorrhizal (VAM) fungi on the seedling growth of three *Pistacia* species. *Journal of Environmental Biology*. 2006;27(3):485-489.
  35. Jamaluddin, Shukla R. Effect of root exudates on colonization of AM fungi in *Jatropha*; 2012.
  36. Muthukumar T, Kathikeyan A, Udaiyan K. Evaluation of the growth and nutrient uptake of *Casuarina equisetifolia* on alfisol soil inoculated with different arbuscular mycorrhizal fungi. In: Jayaraj, R.S.C., Warriar, R.R., Nicodemus, A. and Krishnakumar, N. (Eds.). *Advances in Casuarina Research in India: Proceedings*

- of the 2<sup>nd</sup> National Seminar on Casuarina. Institute of Forest Genetics and Tree Breeding, Coimbatore, India. 2012;49-58.
37. Mohan V, Sandeep C. Biochemical analysis and growth enhancement studies of important medicinal plant, *Rauvolfia serpentina* inoculated with *Arbuscular mycorrhiza* fungi in nursery. International Journal of Current Microbiology and Applied Sciences. 2015;4(6):811- 820.
38. Ortiz YR, Bello CHÁ, Alarcón A, Cerrato RF. Effectiveness of native arbuscular mycorrhiza on the growth of four tree forest species from the Santa Marta Mountain, Veracruz (Mexico). Forest Systems. 2017;26(1):3.

© 2022 Gulab et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*

*The peer review history for this paper can be accessed here:*  
<https://www.sdiarticle5.com/review-history/85299>