



Identification and Isolation of Fungi in Abattoir and Poultry Amended Plots in Ilorin, Southern Guinea Savanna

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Author's contributions

This work was carried out in collaboration between all authors. Author MAA designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors ARS and IA managed the analyses of the study, while authors MI and AOA managed the literature searches. All authors read and approved the final manuscript.

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ABSTRACT

Traditional methods for soil remediation are often expensive and energy consuming and this has given rise to a new and ecologically safer method known as mycoremediation. A field experiment was carried out at the University of Ilorin Dam site to isolate and identify fungi present in organic amended plots which are capable of remediating heavy metal polluted soil of Asa River. Randomized Complete Block design in split plot arrangement was adopted using two treatments: Poultry Dropping (PD) and Abattoir Effluent (AE) at five levels i.e control, 1.3 t/ha, 2.6 t/ha, 1.3 t/ha+NPK120KgN, 2.6t/ha+NPK120KgN for abattoir effluent, and poultry droppings at control, 10t/ha, 15t/ha, 10t/ha+NPK120KgN and 15 t/ha+NPK120KgN having three (3) replicates. Soil samples collected before and after planting were analysed for heavy metals (Mn, Fe, Pb, Zn, Cu, Co, Ni, Cr, Cd) using Atomic Adsorption Spectrophotometer. Result obtained showed that organic

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waste are effective in bioremediation of Asa River sediment with abattoir effluent having a reduction of 99.04% and poultry dropping 98.72% of heavy metal concentration in the soil which varied in the order of: Mn>Fe>Pb>Zn>Cu>Co>Ni>Cr>Cd. Result obtained also showed that eleven metal resistant Fungi were identified from abattoir effluent and poultry dropping amended plots i.e *Aspergillus niger*, *Trichoderma viride*, *Fusarium solani*, *Penicillium notatum*, *Aspergillus flavus*, *Trichoderma harzianum*, *Trichophyton verrucosum*, *Fusarium oxysporum*, *Stachybotrys chartarum*, *Aspergillus ustus* and *Microsporum nanum*. *Aspergillus niger* was observed to have the highest population (19% in poultry droppings and 19.6% in abattoir effluent).

Keywords: *Microremediation; abattoir effluent; fungi; heavy metal and Asa river.*

1. INTRODUCTION

Industrialization, global increase in population and indiscriminate disposal of waste has become a serious concern due to their release of heavy metals which has negative effect on the environment. Several toxic metals from industrial wastewater and other human activities are directly or indirectly released into the environment. Unlike organic contaminants, these pollutants from heavy metals are not biodegradable and able to travel up the food chain via bioaccumulation [1].

Several conventional methods have been used for the removal and treatment of heavy metals in contaminated sites which include electrochemical treatment, ion exchange, precipitation, reverse osmosis, evaporation and sorption [2]. However, these methods are uneconomical due to high reagents and energy requirements, or ineffective at removing all metal contamination, and they can generate large quantities of toxic sludge [3].

More importantly, bioremediation offers a safer and more economical option for treating heavy metals in contaminated sites [4]. The use of microorganisms and organic waste have several advantages, not least the highly efficient removal of heavy metals and cost effectiveness [5].

Some fungi are known to be particularly tolerant of heavy metals [6,7], and many species can adapt and grow under conditions of extreme pH, temperature, nutrient availability and high metal concentrations [8]. Fungi tolerate and detoxify metals through various mechanisms involving valence transformation, extra- and intracellular precipitation or active uptake [9,10]. Scientists are currently exploring the use of microbes and associated biota residing in ecosystems for the bioremediation of pollutants through degradation or accumulation [11], and numerous strains isolated from contaminated sites possess such abilities.

Fungal biomass proves to be advantageous in having a high percentage of cell wall materials which offer excellent metal binding properties [12,13]. Fungal cell walls can act as a cation exchanger due to their negative charge originating from the presence of different functional groups, e.g. carboxylic, phosphate, amine or sulfhydryl, in different wall components (hemicelluloses, pectin, lignin, etc.). Cell walls of fungi are rich in polysaccharides and glycoproteins such as glucans, chitin, mannans and phospho-mannans. Metals and their compounds can interact with fungi in various ways depending on the type of metal, organism and environment [14]. Moreover the use of fungal biomasses as biosorption materials is very convenient because of their inexpensive production methods based on simple fermentation techniques [15]. Fungi can also serve as an economical and constant supply of biomass for biosorption of heavy metals and they can also be grown using inexpensive media and unsophisticated fermentation techniques.

Many fungal species such as *Rhizopus arrhizus*, *Penicillium spinulum* and *Aspergillus niger* have been extensively studied for heavy metals biosorption and the process mechanism seems to be dependent upon species [16,17].

Therefore, this research was aimed at using organic waste to bioremediate Asa River Sediment and also to identify the fungi biomass present in the amended plots.

2. MATERIALS AND METHODS

2.1 Site Description

The research was carried out during the dry season of 2015/2016 at the University of Ilorin Dam Site, Ilorin, Kwara State, Nigeria. The site was located in the Southern Guinea Savanna (SGS) belt at longitude N08°28.049' and latitude E004°39.798', approximately 344.7 m above the

sea level. The average temperature is 28°C and an annual rainfall is 1100-1400 mm per annum.

2.2 Experimental Layout

Randomized Complete Block design in split plot arrangement was adopted, using three (3) treatments: Rice Husk (RH), Poultry dropping (PD) and Abattoir Effluent (AE) at five (5) levels i.e control, 10 t/ha, 15 t/ha, 10 t/ha+NPK and 15 t/ha+NPK for rice husk and poultry dropping, while abattoir effluent were applied at control, 1.3 t/ha, 2.6 t/ha, 1.3 t/ha+NPK and 2.6 t/ha+NPK having three (3) replicate. The treatment combination followed a randomized arrangement.

Rice husk was obtained from Rice Milling Market at Iyana Oja Gboro, Ilorin, while abattoir effluent was collected from Ilorin Abattoir Center, Iyata market, Ilorin and poultry dropping from Unilorin Poultry Unit at the Teaching and Research Farm, Ilorin, Kwara State. All samples were collected in clean sacs and plastic containers as appropriate prior to analysis and field incorporation.

The area was mechanically ploughed and marking of the experimental plot was done manually. The treatments were spread uniformly over the plots and incorporated manually into the seedbed. One seed of Low Nitrogen Tolerant population white (LNTPw) maize (Early maturing variety) were planted per hill at a spacing of 25 cm x 75 cm and depth of 2 - 5 cm fourteen days after incorporation of amendments. The experiment was carried out in dry season and water was supplied through irrigation at an interval of 3 days.

2.3 Collection of Soil Sample

Soil samples were collected from six points on the field to form a composite sample and used to determine soil physical and chemical properties of the soil before amendment was applied. Soil samples were also taken from three points on each plot at a depth of 0 – 30 cm after harvest and bulked together to form composite samples. Soil samples were collected using soil auger.

2.4 Chemical Analysis of Sediment

The pH was determined by the method outlined by [18] using an electronically Jenway 3015 pH meter at ratio of 1:2.5 soil/water. Exchangeable acidity of the soil was determined by titration method using 1N KCl extract as described by [19]. Exchangeable cations were extracted with

an excess of 1 M NH₄OAc (ammonium acetate) where sodium and potassium were determined using flame photometry, and atomic absorption spectrophotometer was used to determine the calcium and magnesium [20]. Organic carbon in sediment was determined using wet oxidation method of [21], Available phosphorus was determined using Bray 1 [22] method. Total Nitrogen was determined using micro-Kjeldhal distillation method by [23].

2.5 Heavy Metals Analysis of Soil

Heavy metals in soil (5 g) was extracted with 1% EDTA as described by [24]. Solutions were filtered through Whatman No. 1 filter paper and volume was made to 50ml by adding distilled water. Soil samples were digested in triplicates and analyzed for Fe, Mn, Zn, Co, Cd, Cr, Cu, Ni and Pb. For the determination of heavy metals the atomic absorption spectrophotometer was powered on and warmed up for 30 minutes. After the heating of cathode lamp, the air acetylene flame was ignited and instrument was calibrated or standardized with different working standards.

2.6 Sterilization of Apparatus

Petri plates, media bottles, distilled water, McCartney bottles and syringes were sterilized in autoclave. For sterilization purpose all apparatus were autoclaved for 40 minutes at 121°C. After autoclaving all sterilized material dried in oven at 95°C.

2.7 Media Preparation and Fungal Isolation

Potato Dextrose Agar (PDA) media was used for the isolation of fungi. Fungi were isolated on Potato Dextrose Agar (PDA) by soil dilution method [33].

2.8 Preparation of Plates

Fungi were isolated on Potato Dextrose Agar (PDA) by soil dilution method [25]. Poured the media in Petri-dishes and allowed to solidify for 24 hour. To suppress the bacterial growth, 30 mg/lit of streptomycin was added in the medium [25]. Once the agar was solidified, and then put plates in an inverted position for 24 hours at room temperature [25].

2.9 Identification of Fungi

The fungal cultures were identified on the basis of macroscopic (colonial morphology, color,

texture, shape, diameter and appearance of colony) and microscopic (septation in mycelium, presence of specific reproductive structures, shape and structure of conidia, and presence of sterile mycelium) characteristics [26]. Pure cultures of fungi isolates were identified with the help of literature [27,28].

3. RESULTS AND DISCUSSION

The Table 1 shows the initial soil chemical properties of Asa River sediment, in which the sediments were observed to be sandy. Organic matter content of the sediment was ranked low at (0.087%), while the nitrogen content of the sediment also ranked as low (0.07). The pH of the sediment showed 8.9 in KCl which means that the sediment was alkaline in nature and available phosphorus was also seen to be extremely low (0.046). Calcium was observed to be low, magnesium and potassium were also observed to be extremely low and sodium also observed to have low value (0.11). Cation exchange capacity of the sediment was observed to fall below the range considered to be low. The chemical analysis of the sediment showed that the soil is low in terms of fertility. The heavy metal analysis of the sediment

revealed that all metals fell below maximum permissible limit as set by USEPA except manganese (350.5 mg/kg). Table 1 also revealed that abattoir effluent has a high nitrogen content of (1.06) compared to poultry dropping of 0.71%. Table 1 further revealed that abattoir effluent had a higher phosphorus content (24.52%) than poultry dropping (11.92%), and potassium, calcium magnesium and sodium were observed to be higher in poultry dropping than abattoir effluent.

Table 2 shows that there were no significant differences at <0.05 between the treatments at various levels of application except for Fe with the highest reduction in concentration observed at sole application rate of 3.3 t/ha. Although they were no significant differences between the treatments applied yet it was observed that the organic waste was very effective in reducing the concentration of heavy metals that were available in the soil. This result is supported by the studies of [29,30] that organic waste possesses large surface area, which implied a high capacity for complexing heavy metals on their surface. Impact of organic amendments in bioremediation is effective and their effectiveness is related to the quantity applied [31].

Table 1. Soil chemical analysis prior to planting

Chemical Properties	Soil	Abattoir effluent	Poultry dropping
Nitrogen%	0.07	1.06	0.71
Available P (mg/kg)	0.046	24.52	11.92
Exchangeable Ca (Cmol/kg)	2.79	11.58	33.87
Exchangeable Mg (Cmol/kg)	0.14	2.61	17.85
Exchangeable K (Cmol/kg)	0.02	4.55	16.93
Exchangeable Na (Cmol/kg)	0.11	0.11	0.14
Organic carbon%	0.5		
Organic Matter%	0.87		
pH in H ₂ O(1:2.5)	9.3		
pH in KCl(1:2.5)	8.9		
Acidity (mol/kg)	0.4		
Sand	75.96		
Silt	11.28		
Clay	12.76		
Textural Class	Sand		
Cation exchange capacity	3.06		
Chromium (mg/kg)	25.3		
Cadmium (mg/kg)	0.5		
Nickel (mg/kg)	2.5		
Lead (mg/kg)	9.1		
Manganese (mg/kg)	350.5		
Iron (mg/kg)	25,250		
Copper (mg/kg)	9.65		
Zinc (mg/kg)	54.7		
Cobalt (mg/kg)	1.9		

Table 2. Effect of organic waste on heavy metal availability in soil after cropping

Treat./Levels	Co (mg/kg)	Mn (mg/kg)	Cr (mg/kg)	Cd (mg/kg)	Pb (mg/kg)	Zn (mg/kg)	Fe (mg/kg)	Cu (mg/kg)	Ni (mg/kg)
Control	0.30	58.1	0.04	0.00	1.8	0.45	4.23	0.47	0.10
AE 1.6 t/ha	0.18	46.9	0.05	0.01	1.53	0.57	3.00	0.48	0.067
AE 3.2 t/ha	0.24	44.0	0.04	0.01	1.47	0.59	2.37	0.51	0.03
AE 1.6 t/ha+NPK120 KgN	0.34	57.9	0.04	0.003	1.5	0.51	4.33	0.45	0.10
AE 3.2 t/ha +NPK120 KgN	0.31	52.2	0.12	0.003	1.7	0.52	4.20	0.38	0.00
Control	0.30	58.1	0.04	0.00	1.8	0.45	4.23	0.47	0.10
PD10 t/ha	0.24	55.6	0.04	0.01	1.5	0.82	2.60	0.55	0.067
PD15 t/ha	0.31	47.0	0.07	0.003	1.77	0.86	3.73	0.62	0.067
PD10 t/ha+NPK120 KgN	0.33	48.0	0.05	0.003	1.37	0.81	2.93	0.50	0.067
PD15 t/ha+NPK120 KgN	0.24	40.5	0.04	0.00	1.37	0.78	4.13	0.42	0.067
SED	0.003	5.32	0.027	0.003	0.246	0.099	0.507	0.07	0.037
LSD(0.05)	ns	ns	ns	ns	ns	ns	*	ns	ns

3.1 Occurrence of Fungi Isolates in Poultry Droppings

Fig. 1 shows the percentage of occurrence of the fungi in poultry dropping amended plots. Ten (10) fungal species were isolated. It shows that *Aspergillus niger* had the highest occurrence (19.1%), followed by *Aspergillus flavus* (14.6%), *Trichoderma viride* (14%), *Trichoderma harzianum* (12.3%), *Fusarium solani* (12.1%), *Penicillium notatum* (10.3), *Trichoderma verrucosum* (6.4%), *Fusarium oxysporum* (5.3%), *Microsporium nanum* (3.8%) and *Aspergillus ustus* (2.1%).

3.2 Occurrence of Fungi Isolates in Abattoir Effluent

Fig. 2 shows the percentage of occurrence of fungi in abattoir effluent. Eleven (11) fungal species were isolated. *Aspergillus niger* had the highest percentage of occurrence (19.6%), followed by *Trichoderma viride* (15.1%), *Fusarium solani* (11.2%), *Penicillium notatum* (10.1%), *Aspergillus flavus* (9.7%), *Trichoderma harzianus* (7.8%), *Trichophyton verrucosum* (6.5%), *Fusarium oxysporum* (6.2%), *Stachybotrys chartarum* (5.4%), *Aspergillus*

ustus (4.6%) while *Microsporium nanum* had the lowest occurrence (3.8%).

Result shows that different fungi of Ascomycotina were isolated and identified in the amended plots and these fungi showed excellent bioremediation capability for different heavy metals such as Co, Cd, Cu, Ni, Pb and Zn. These agrees with the findings of [32] who evaluated the efficiency of different fungi and found out that *Aspergillus niger* and *A. flavus* has high capacity in remediating Pb, Zn, Cr and Cd from refinery effluent. Furthermore, [33] in their study of Mycoremediation: utilization of fungi for reclamation of heavy metals at their optimum remediation conditions found out that *Aspergillus niger*, *A. flavus*, *A. fumigates* and *A. ustus* had great potential in reducing heavy metals concentration in contaminated environment. This finding was similar to that of [34,35]. Two *Fusarium* sp. were identified in which *Fusarium solani* was known to degrade Cr, Zn and Nickel [36], while *F. oxysporum* was also identified to be effective in reducing the concentration of Cd, Cu, Pb, and Zn. This is similar to the findings of [37] who discovered the *F. oxysporum* helps to biosorb Zn and Cu and *F. solani* removes Cu [38]. Similarly, two species of *Trichoderma* were identified; *T. harzianum* and *T. viride*. *T. viride*

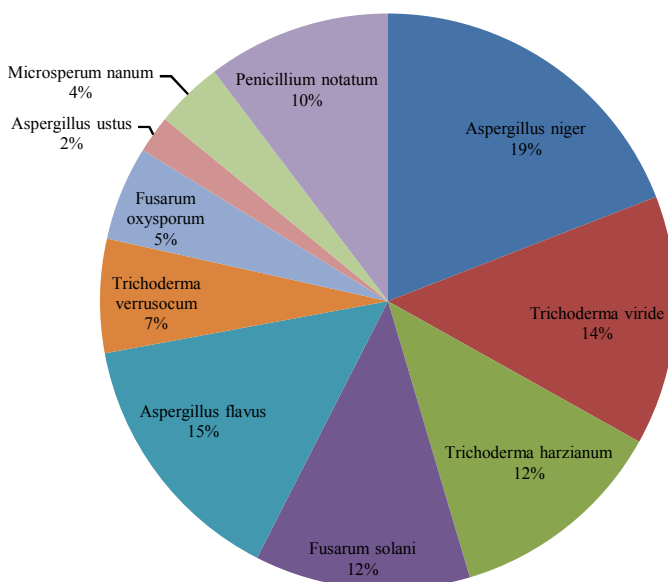


Fig. 1. Percentage occurrence of fungi in poultry droppings

was identified to have a high accumulating potential for Ni, Cd and Cr [39], and [40] found out that *T. viride* has high uptake potential for

chromium. While *T. harzanium* has been observed to have a high reducing capacity for Siddique et al., 2011, [41]. Moreso, *P. notatum*

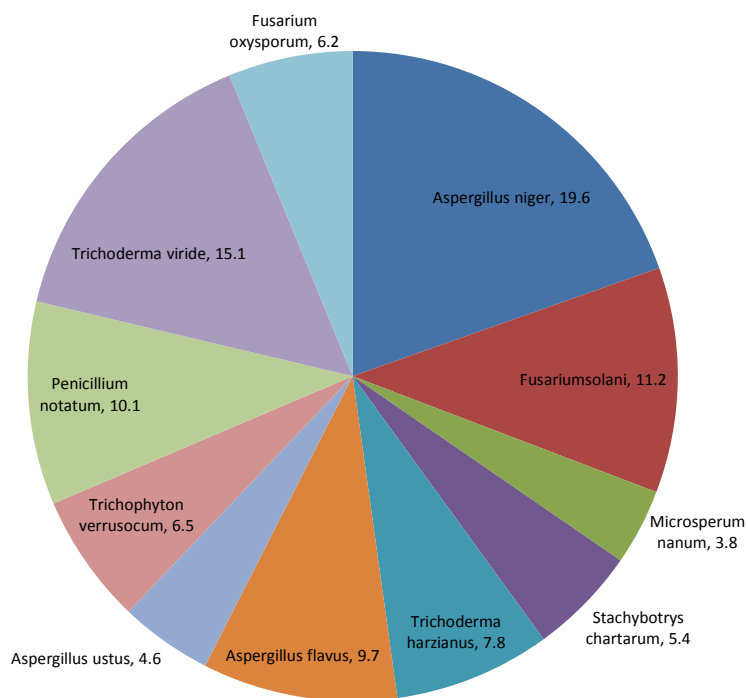


Fig. 2. Percentage occurrence of fungi in abattoir effluent

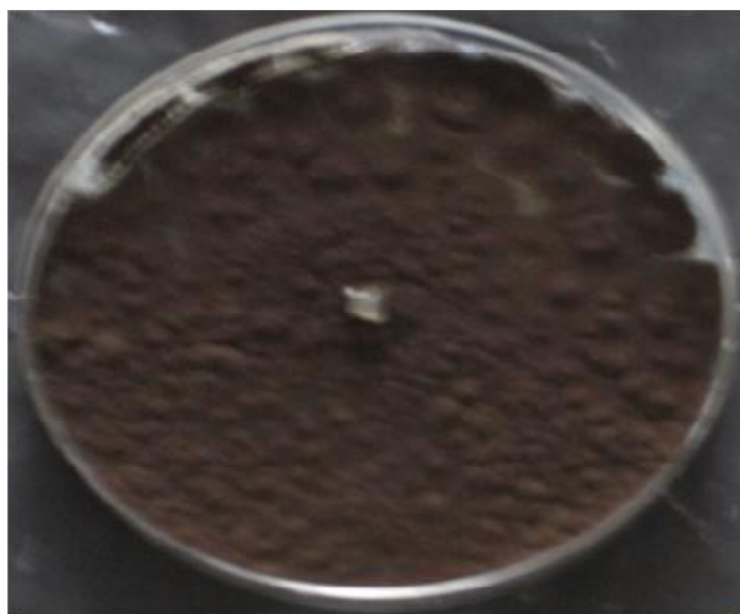


Plate 1(a). Colony morphology of *Aspergillus niger*



Plate 1(b). Microscopic structure of *Aspergillus niger* X100

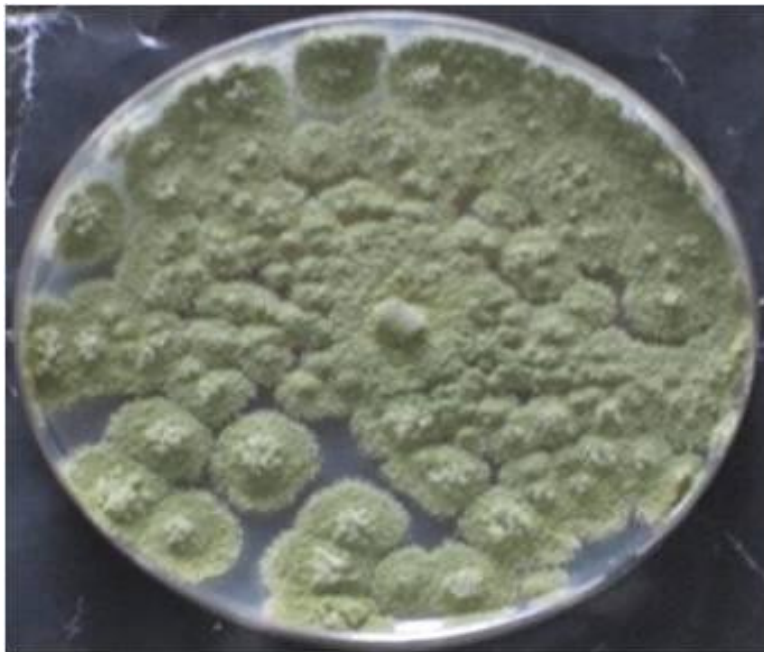


Plate 2(a). Colony morphology of *Aspergillus flavus*

was found to have capacity for reducing high chromium concentrations [42]. Result further revealed that *Stachybotrys chartarum*, played an important role in reducing copper (Cu) in contaminated soil. In a study conducted by [43] to evaluate the tolerance capacity of *Stachybotrys chartarum* in a copper based

preservative, they found that the fungi had high tolerance level for copper. Furthermore, *Microsporum nanum* was found to play an important role on Cd, Cr, Pb and this was supported by the findings of [32] who opined that *M. nanum* reduced the concentration of heavy metal from a refinery effluent.



Plate 2(b). Microscopic structure of *Aspergillus flavus* X100



Plate 3(a). Colony morphology of *Fusarium oxysporum*

It was also observed from Figs. 1 and 2 that *Aspergillus niger* was the dominant isolated specie from the two amended plots. *Aspergillus niger* has been known to be able to tolerate the heavy metals in organic wastes and the result

obtained in this study confirm the earlier assertion by different authors who asserted that *Aspergillus sp.* was the dominant species in wastewater [44].

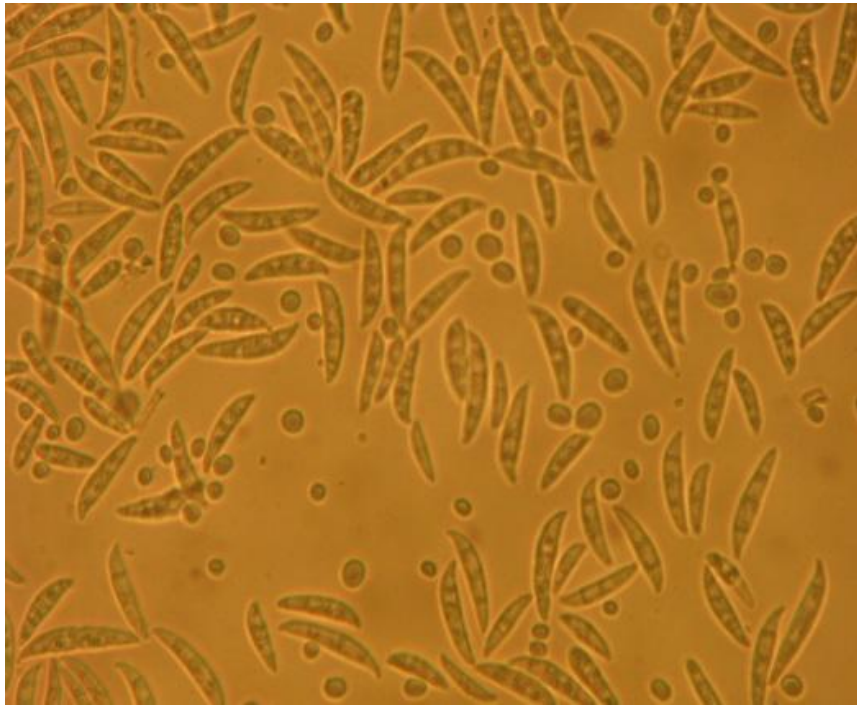


Plate 3(b). Microscopic structure of *Fusarium oxysporum* X 100

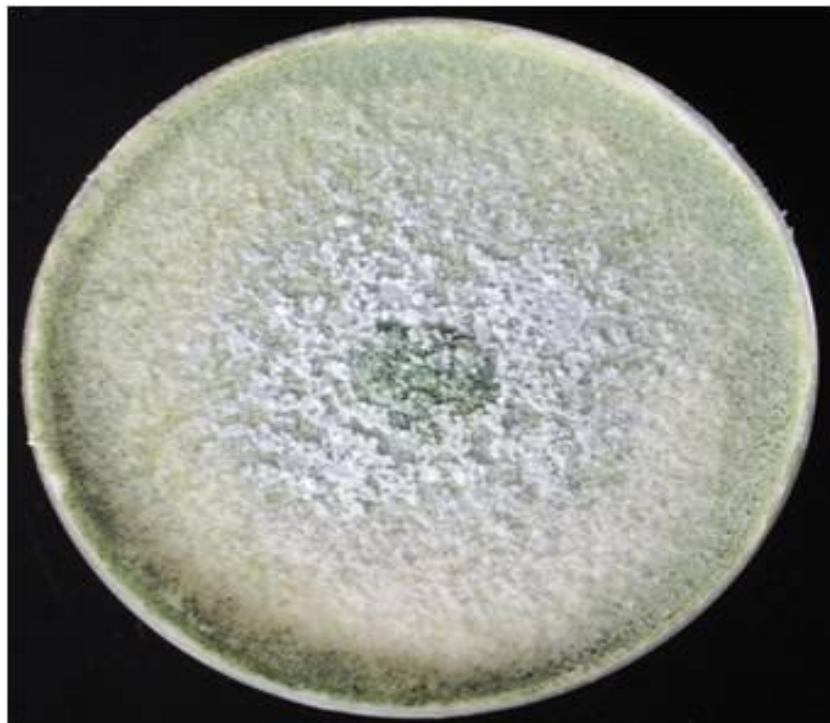


Plate 4(a). Colony morphology of *Trichoderma harzianum*

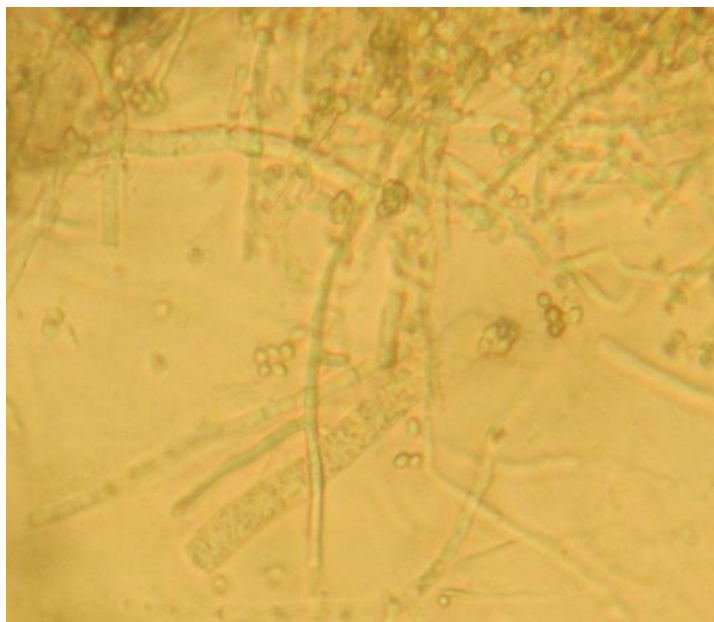


Plate 4(b). Microscopic structure of *Trichoderma harzianum* X100

4. CONCLUSION

From the result of this study, it was deduced that organic wastes are effective material for bioremediation, and their effectiveness was enhanced as a result of the activities of certain fungi which performed important functions coupled with their inherent potential to tolerate and reduce heavy metals. Fungi isolated and identified in abattoir and poultry amended plots includes *Aspergillus niger*, *Trichoderma viride*, *Fusarium solani*, *Penicillium notatum*, *Aspergillus flavus*, *Trichoderma harzianum*, *Trichophyton verrucosum*, *Fusarium oxysporum*, *Stachybotrys chartarum*, *Aspergillus ustus* and *Microsporium nanum*. *Aspergillus niger* (19% in poultry droppings and 19.6% in abattoir effluent) was observed to have the highest population in all the amended plots.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Rovina K, Azad SA, Naher L, Suryani S, Siddiquee S. Heavy metal contaminants removal from wastewater using the potential filamentous fungi biomass. Journal of Microbial & Biochemical Technology Journal of Microbial & Bioch. Technol. 2015;7:384-393.
2. Kadirvelu K, Senthilkumar P, Thamaraiselvi K, Subburam V. Activated carbon prepared from biomass as adsorbent: Elimination of Ni(II) from aqueous solution. Bioresour. Technol. 2002;81:87-90.
3. Hemambika B, Johncy RM, Rajesh VK. Biosorption of heavy metals by immobilized and dead fungal cells: A comparative assessment. Journal of Ecology and the Natural Environment Vol. 2011;3(5):168-175.
4. Iskandar NL, Zainudin NAIM, Tan SG. Tolerance and biosorption of copper (Cu) and lead (Pb) by filamentous fungi isolated from a freshwater ecosystem. Journal of Environmental Science. 2011;23(5):824-830.
5. Concas A, Cao G. Technologies for the Remediation of Contaminated Sites (Italian Language), CUEC. 2004;1-85.
6. Gavrilesca M, Removal of heavy metals from the environment by biosorption. Eng. Life Sci. 2004;4(3):219-232.
7. Baldrian P. Interactions of heavy metals with white-rot fungi. Enzyme Microb. Technol. 2003;32:78-83.
8. Isar P, Saran JS, Saxena RK, Bioaccumulation of copper by *Trichoderma viride*. Bioresour. Technol. 2006;97:1018-1021.

9. Mala JGS, Nair BU, Puanakrishnan R. Accumulation and biosorption of chromium by *Aspergillus niger* MTCC 2594. J. Gen. Appl. Microbiol. 2006;52:179-186.
10. Turnau K, Orlowska E, Ryszka P, Zubek S, Anielska T, Gawronski S, Jurkiewicz A. Role of mycorrhizal fungi in phytoremediation and toxicity monitoring of heavy metal rich industrial wastes in southern Poland. Soil Water Pollut. Monit. Prot. Remed. 2006;3(23):533-551.
11. Cao SQ, Zheng YM, Huang YZ, Zhu YG. Health risks of heavy metals in contaminated soils and food crops irrigated with wastewater in Beijing, China. Environ. Pollut. 2008;152:686-692.
12. Mann H. Biosorption of heavy metals by bacterial biomass. In: Volesky B (ed) Biosorption of heavy metals. CRC Press, Boca Raton, FL. 1990;93-137.
13. Luef E, Prey T, Kubicek CP. Biosorption of Zinc by fungal mycelia waste. Appl. Microbiol. Biotechnol. 1991;34:688-692.
14. Aung MM, Ing YP. Bioleaching of spent fluid catalytic cracking catalyst using *Aspergillus niger*, Journal of Biotechnology, 2005;116:159-170.
15. Maurya NS, Mittal AK, Cornel P, Rother E. Biosorption of dyes using dead macro fungi: Effect of ye structure, ionic strength and pH, Bioresour. Technol. 2006;97:512-521.
16. Hafez N, Abdel-Razek AS, Hafez MB. Accumulation of some heavy metals on *Aspergillus flavus*. J Chem Technol Biotechnol. 1997;68:19-22.
17. Kapoor A, Viraraghavan T. Heavy metal biosorption sites in *Aspergillus niger*. Biores Technol. 1997;61:221-227
18. Bates RG. *Electromeric pH determination*. John Wiley and sons Inc., New York, USA. 1954;87-92.
19. Rhoades JD. Cation exchange capacity. In: AL. Page (ed.) Methods of soil analysis. Part 2: Chemical and microbiological properties (2nd ed.) Agronomy. 1982;9: 149-157.
20. Anderson JM, Ingram JSI. Tropical Soil Biology and Fertility: A Handbook of Methods. Second edition. CAB International, Wallingford, UK. 1993;37.
21. Walkley A, and Black TA. An examination of the Degtjarett methods for determining soil organic matter and a proposed modification of the chromic acid titration method. Journal of Soil Science. 1934;37: 29-38.
22. Bray RH, Kurtz LT, Determination of total, organic, and available forms of phosphorus in soils. Soil Science. 1945;59:39-45.
23. AOAC International (formerly the Association of Official Analytical Chemists). Official Methods of Analysis. Arlington, VA: AOAC International; 1999.
24. Association of official analytical chemist. Official methods of analysis. 13th edition. AOAC, Washington, DC; 1980.
25. Razak AA, Bachman G, Farrag R. Activities of microflora in soils of upper and lower Egypt. African Journal of Mycology and Biotechnology. 1999;7:1-19.
26. Zafar S, Aqil F, Ahmad I. Metal tolerance and biosorption potential of filamentous fungi isolated from metal contaminated agriculture soil. Bioresource Technology. 2007;98:2557-2561.
27. Domsch KH, Gams W, Anderson TH. Compendium of soil fungi. London, England: Academic Press. 1980
28. Barnett HL, Hunter BB. Illustrated genera of imperfect fungi, fourth edition. Prentice Hall Inc.
29. Uchimiya M, Lima IM, Klasson KT, Wartelle LH. Contaminant immobilization and nutrient release by biochar soil amendment. Roles of Natural Organic Matter, Chemosphere. 2010;80: 935-940.
30. Park JH, Choppala GH, Bolan NS, Chung JW, Chuasavathi T. Biochar reduces the bioavailability and phytotoxicity of heavy metals, Plant Soil. 2011;348:439-451.
31. Walker DJR, Clemente A, Roig MP, Bernal, 2003. The effects of soil amendments on heavy metal bioavailability in two contaminated Mediterranean soils. Environ. Pollut. 2003; 122:303-312.
32. Bello OA, Abubakar BY, Abdullahi IO. Efficiency of *Aspergillus niger*, *Aspergillus flavus* and *Microsporium nanum* to remove heavy metal from refinery effluent. Journal of Advances in Biology and Biotechnology, 2016;6(3):1-6.
33. Archana, Jaitly AK. Mycoremediation: utilization of fungi for reclamation of heavy metals at their optimum remediation conditions. Journal of Biology and Life Sciences. 2015; 3(1):77-106.
34. Ahmad I, Ansari I, Aqil F. Biosorption of Ni, Cr and Cd by metal tolerant *Aspergillus niger* and *Penicillium sp.* using single and

- multi-metal solution. Indian J Exp. Biol. 2005a;44:73-76.
35. Ramasamy RK, Congeevaram S, Thamaraiselvi K. Evaluation of isolated fungal strain from e-waste recycling facility for effective sorption of toxic heavy metal pb (II) ions and fungal protein molecular characterization- a mycoremediation approach. Asian J. Exp. Biol. Sci. 2011;2: 342-247.
36. Sen M. Biosorption of zinc and nickel from wastewaters using nonliving cells of *Fusarium solani*. Int. J. Chem. Tech. Appl. 2013;2:63-70.
37. Chandrakar V, Verma P, Jamaluddin. Removal of cu and zn by fungi in municipal sewage water. Int. J. Adv. Biotechnol. Res. 2012;2:787-790.
38. Amatussalam A, Abubacker MN, Rajendran RB. *In situ* Carica papaya stem matrix and *Fusarium oxysporum* (NCBT-156) mediated bioremediation of chromium. Indian J Exp. Biol. 2011;49: 925-31.
39. Levinskaite L. Simultaneous effect of Ni, Cd and Cr on soil micromycetes. Biologija. 2001;13-15.
40. Joshi PK, Swarup A, Maheshwari S, Kumar R, Singh N. Bioremediation of heavy metals in liquid media through fungi isolated from contaminated sources. Indian J. Microbial. 2011;51:482-487.
41. Sarkar S, Satheshkumar A, Jayanthi R, Premkumar R. Biosorption of Nickel by live biomass of *Trichoderma harzianum*. Res. J. Agr. Sci. 2001;1:69-74.
42. Seshikala D, Charya MAS. Effect of pH on Chromium biosorption. Int. J. Pharma. Bio. Sci. 2012;2:298-302.
43. Danguolé B, Loreta L. Fungal tolerance towards copper-based wood preservatives. BIOLOGIJA. 2007;53(4):54-61.
44. Sharma Kavita. Isolation of soil mycoflora of katao near Gangtok, India. Journal of Phytology. 2010;2(5):30-32.

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